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(54) Title: ANTISENSE OLIGONUCLEOTIDE MODULATION OF *raf* GENE EXPRESSION

(57) Abstract

Oligonucleotides are provided which are targeted to nucleic acids encoding human raf and capable of inhibiting raf expression. In preferred embodiments, the oligonucleotides are targeted to mRNA encoding human c-raf or human A-raf. The oligonucleotides may have chemical modifications at one or more positions and may be chimeric oligonucleotides. Methods of inhibiting the expression of human raf using oligonucleotides of the invention are also provided. The present invention further comprises methods of inhibiting hyperproliferation of cells and methods of treating abnormal proliferative conditions which employ oligonucleotides of the invention.

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ANTISENSE OLIGONUCLEOTIDE MODULATION OF raf GENE EXPRESSION

FIELD OF THE INVENTION

This invention relates to compositions and methods for modulating expression of the raf gene, a naturally present cellular gene which has been implicated in abnormal cell proliferation and tumor formation. This invention is also directed to methods for inhibiting hyperproliferation of cells; these methods can be used diagnostically or therapeutically. Furthermore, this invention is directed to treatment of conditions associated with expression of the raf gene.

BACKGROUND OF THE INVENTION

Alterations in the cellular genes which directly or indirectly control cell growth and differentiation are considered to be the main cause of cancer. The raf gene family includes three highly conserved genes termed A-, B- and c-raf (also called raf-1). Raf genes encode protein kinases that are thought to play important regulatory roles in signal transduction processes that regulate cell proliferation. Expression of the c-raf protein is believed to play a role in abnormal cell proliferation since it has been reported that 60% of all lung carcinoma cell lines express unusually high levels of c-raf mRNA and protein. Rapp et al., *The Oncogene Handbook*, E.P. Reddy, A.M Skalka and T. Curran, eds., Elsevier Science Publishers, New York, 1988, pp. 213-253.

Oligonucleotides have been employed as therapeutic moieties in the treatment of disease states in animals and man. For example, workers in the field have now identified antisense, triplex and other oligonucleotide compositions which are capable of modulating expression of genes implicated in viral, fungal and metabolic diseases.

As examples, U. S. Patent 5,135,917, issued August 4, 1992, provides antisense oligonucleotides that inhibit human interleukin-1 receptor expression. U.S. Patent 5,098,890, issued March 24, 1992 in the name of Gewirtz et al., is

directed to antisense oligonucleotides complementary to the c-myb oncogene and antisense oligonucleotide therapies for certain cancerous conditions. U.S. Patent 5,087,617, issued February 11, 1992, provides methods for treating cancer patients with antisense oligonucleotides. U.S. Patent 5,166,195 issued November 24, 1992, provides oligonucleotide inhibitors of HIV. U.S. Patent 5,004,810, issued April 2, 1991, provides oligomers capable of hybridizing to herpes simplex virus Vmw65 mRNA and inhibiting replication. U.S. Patent 5,194,428, issued March 16, 1993, provides antisense oligonucleotides having antiviral activity against influenzavirus. U.S. Patent 4,806,463, issued February 21, 1989, provides antisense oligonucleotides and methods using them to inhibit HTLV-III replication. U.S. Patent 5,286,717 (Cohen et al.), issued February 15, 1994, is directed to a mixed linkage oligonucleotide phosphorothioates complementary to an oncogene; U.S. Patent 5,276,019 and U.S. Patent 5,264,423 (Cohen et al.) are directed to phosphorothioate oligonucleotide analogs used to prevent replication of foreign nucleic acids in cells. Antisense oligonucleotides have been safely administered to humans and clinical trials of several antisense oligonucleotide drugs, targeted both to viral and cellular gene products, are presently underway. The phosphorothioate oligonucleotide, ISIS 2922, has been shown to be effective against cytomegalovirus retinitis in AIDS patients. *BioWorld Today*, April 29, 1994, p. 3. It is thus established that oligonucleotides can be useful therapeutic instrumentalities and can be configured to be useful in treatment regimes for treatment of cells and animal subjects, especially humans.

Antisense oligonucleotide inhibition of gene expression has proven to be a useful tool in understanding the roles of raf genes. An antisense oligonucleotide complementary to the first six codons of human c-raf has been used to demonstrate that the mitogenic response of T cells to interleukin-2 (IL-2) requires c-raf. Cells treated with the oligonucleotide showed a near-total loss of c-raf protein and

a substantial reduction in proliferative response to IL-2. Riedel et al., *Eur. J. Immunol.* **1993**, 23, 3146-3150. Rapp et al. have disclosed expression vectors containing a raf gene in an antisense orientation downstream of a promoter, and methods of inhibiting raf expression by expressing an antisense Raf gene or a mutated Raf gene in a cell. WO application 93/04170. An antisense oligodeoxyribonucleotide complementary to codons 1-6 of murine c-Raf has been used to abolish insulin stimulation of DNA synthesis in the rat hepatoma cell line H4IIE. Tornkvist et al., *J. Biol. Chem.* **1994**, 269, 13919-13921. WO Application 94/06248 discloses methods for identifying an individual at increased risk of developing cancer and for determining a prognosis and proper treatment of patients afflicted with cancer comprising amplifying a region of the c-raf gene and analyzing it for evidence of mutation.

Denner et al. disclose antisense polynucleotides hybridizing to the gene for raf, and processes using them. WO 94/15645. Oligonucleotides hybridizing to human and rat raf sequences are disclosed.

Iversen et al. disclose heterotypic antisense oligonucleotides complementary to raf which are able to kill ras-activated cancer cells, and methods of killing raf-activated cancer cells. Numerous oligonucleotide sequences are disclosed, none of which are actually antisense oligonucleotide sequences.

There remains a long-felt need for improved compositions and methods for inhibiting raf gene expression.

SUMMARY OF THE INVENTION

The present invention provides oligonucleotides which are targeted to nucleic acids encoding human raf and are capable of inhibiting raf expression. The present invention also provides chimeric oligonucleotides targeted to nucleic acids encoding human raf. The oligonucleotides of the invention are believed to be useful both diagnostically and therapeutically, and are believed to be particularly useful in the methods of the present invention.

The present invention also comprises methods of inhibiting the expression of human raf, particularly the abnormal expression of raf. These methods are believed to be useful both therapeutically and diagnostically as a consequence of the association between raf expression and hyperproliferation. These methods are also useful as tools, for example for detecting and determining the role of raf expression in various cell functions and physiological processes and conditions and for diagnosing conditions associated with raf expression.

The present invention also comprises methods of inhibiting hyperproliferation of cells using oligonucleotides of the invention. These methods are believed to be useful, for example in diagnosing raf-associated cell hyperproliferation. Methods of treating abnormal proliferative conditions are also provided. These methods employ the oligonucleotides of the invention. These methods are believed to be useful both therapeutically and as clinical research and diagnostic tools.

DESCRIPTION OF THE DRAWING

Figure 1 is a line graph showing the effect of ISIS 5132 (Figure 1A) and a scrambled control oligonucleotide ISIS 10353 (Figure 1B) on growth of A549 lung tumor xenografts in nude mice. ISIS 5132 decreased tumor size at all doses (0.006 mg/kg; 0.06 mg/kg; 0.6 mg/kg; and 6.0 mg/kg) in a dose-dependent manner. The scrambled raf oligonucleotide, ISIS 10353, had no effect at any dose (Figure 1B).

DETAILED DESCRIPTION OF THE INVENTION

Malignant tumors develop through a series of stepwise, progressive changes that lead to the loss of growth control characteristic of cancer cells, i.e., continuous unregulated proliferation, the ability to invade surrounding tissues, and the ability to metastasize to different organ sites. Carefully controlled *in vitro* studies have helped define the factors that characterize the growth of normal and neoplastic cells and have led to the identification of specific proteins that control

cell growth and differentiation. The raf genes are members of a gene family which encode related proteins termed A-, B- and c-raf. Raf genes code for highly conserved serine-threonine-specific protein kinases. These enzymes are differentially expressed; c-raf, the most thoroughly characterized, is expressed in all organs and in all cell lines that have been examined. A- and B-raf are expressed in urogenital and brain tissues, respectively. c-raf protein kinase activity and subcellular distribution are regulated by mitogens via phosphorylation. Various growth factors, including epidermal growth factor, acidic fibroblast growth factor, platelet-derived growth factor, insulin, granulocyte-macrophage colony-stimulating factor, interleukin-2, interleukin-3 and erythropoietin, have been shown to induce phosphorylation of c-raf. Thus, c-raf is believed to play a fundamental role in the normal cellular signal transduction pathway, coupling a multitude of growth factors to their net effect, cellular proliferation.

Certain abnormal proliferative conditions are believed to be associated with raf expression and are, therefore, believed to be responsive to inhibition of raf expression. Abnormally high levels of expression of the raf protein are also implicated in transformation and abnormal cell proliferation. These abnormal proliferative conditions are also believed to be responsive to inhibition of raf expression. Examples of abnormal proliferative conditions are hyperproliferative disorders such as cancers, tumors, hyperplasias, pulmonary fibrosis, angiogenesis, psoriasis, atherosclerosis and smooth muscle cell proliferation in the blood vessels, such as stenosis or restenosis following angioplasty. The cellular signalling pathway of which raf is a part has also been implicated in inflammatory disorders characterized by T-cell proliferation (T-cell activation and growth), such as tissue graft rejection, endotoxin shock, and glomerular nephritis, for example.

It has now been found that elimination or reduction of raf gene expression may halt or reverse abnormal cell

proliferation. This has been found even in when levels of raf expression are not abnormally high. There is a great desire to provide compositions of matter which can modulate the expression of the raf gene. It is greatly desired to provide methods of detection of the raf gene in cells, tissues and animals. It is also desired to provide methods of diagnosis and treatment of abnormal proliferative conditions associated with abnormal raf gene expression. In addition, kits and reagents for detection and study of the raf gene are desired.

"Abnormal" raf gene expression is defined herein as abnormally high levels of expression of the raf protein, or any level of raf expression in an abnormal proliferative condition or state.

The present invention employs oligonucleotides targeted to nucleic acids encoding raf. This relationship between an oligonucleotide and its complementary nucleic acid target to which it hybridizes is commonly referred to as "antisense". "Targeting" an oligonucleotide to a chosen nucleic acid target, in the context of this invention, is a multistep process. The process usually begins with identifying a nucleic acid sequence whose function is to be modulated. This may be, as examples, a cellular gene (or mRNA made from the gene) whose expression is associated with a particular disease state, or a foreign nucleic acid from an infectious agent. In the present invention, the target is a nucleic acid encoding raf; in other words, the raf gene or mRNA expressed from the raf gene. The targeting process also includes determination of a site or sites within the nucleic acid sequence for the oligonucleotide interaction to occur such that the desired effect- modulation of gene expression- will result. Once the target site or sites have been identified, oligonucleotides are chosen which are sufficiently complementary to the target, i.e., hybridize sufficiently well and with sufficient specificity, to give the desired modulation.

In the context of this invention "modulation" means either inhibition or stimulation. Inhibition of raf gene expression is presently the preferred form of modulation. This

modulation can be measured in ways which are routine in the art, for example by Northern blot assay of mRNA expression or Western blot assay of protein expression as taught in the examples of the instant application. Effects on cell proliferation or tumor cell growth can also be measured, as taught in the examples of the instant application. "Hybridization", in the context of this invention, means hydrogen bonding, also known as Watson-Crick base pairing, between complementary bases, usually on opposite nucleic acid strands or two regions of a nucleic acid strand. Guanine and cytosine are examples of complementary bases which are known to form three hydrogen bonds between them. Adenine and thymine are examples of complementary bases which form two hydrogen bonds between them. "Specifically hybridizable" and "complementary" are terms which are used to indicate a sufficient degree of complementarity such that stable and specific binding occurs between the DNA or RNA target and the oligonucleotide. It is understood that an oligonucleotide need not be 100% complementary to its target nucleic acid sequence to be specifically hybridizable. An oligonucleotide is specifically hybridizable when binding of the oligonucleotide to the target interferes with the normal function of the target molecule to cause a loss of utility, and there is a sufficient degree of complementarity to avoid non-specific binding of the oligonucleotide to non-target sequences under conditions in which specific binding is desired, i.e., under physiological conditions in the case of *in vivo* assays or therapeutic treatment or, in the case of *in vitro* assays, under conditions in which the assays are conducted.

In preferred embodiments of this invention, oligonucleotides are provided which are targeted to mRNA encoding c-raf and A-raf. In accordance with this invention, persons of ordinary skill in the art will understand that mRNA includes not only the coding region which carries the information to encode a protein using the three letter genetic code, but also associated ribonucleotides which form a region known to such persons as the 5'-untranslated region, the 3'-

untranslated region, the 5' cap region, intron regions and intron/exon or splice junction ribonucleotides. Thus, oligonucleotides may be formulated in accordance with this invention which are targeted wholly or in part to these associated ribonucleotides as well as to the coding ribonucleotides. In preferred embodiments, the oligonucleotide is targeted to a translation initiation site (AUG codon) or sequences in the 5'- or 3'-untranslated region of the human c-raf mRNA. The functions of messenger RNA to be interfered with include all vital functions such as translocation of the RNA to the site for protein translation, actual translation of protein from the RNA, splicing or maturation of the RNA and possibly even independent catalytic activity which may be engaged in by the RNA. The overall effect of such interference with the RNA function is to cause interference with raf protein expression.

The present invention provides oligonucleotides for modulation of raf gene expression. Such oligonucleotides are targeted to nucleic acids encoding raf. Oligonucleotides and methods for modulation of c-raf and A-raf are presently preferred; however, compositions and methods for modulating expression of other forms of raf are also believed to have utility and are comprehended by this invention. As hereinbefore defined, "modulation" means either inhibition or stimulation. Inhibition of raf gene expression is presently the preferred form of modulation.

In the context of this invention, the term "oligonucleotide" refers to an oligomer or polymer of nucleotide or nucleoside monomers consisting of naturally occurring bases, sugars and intersugar (backbone) linkages. The term "oligonucleotide" also includes oligomers comprising non-naturally occurring monomers, or portions thereof, which function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of properties such as, for example, enhanced cellular uptake and increased stability in the presence of nucleases.

Certain preferred oligonucleotides of this invention are chimeric oligonucleotides. "Chimeric oligonuclectides" or

"chimeras", in the context of this invention, are oligonucleotides which contain two or more chemically distinct regions, each made up of at least one nucleotide. These oligonucleotides typically contain at least one region of modified nucleotides that confers one or more beneficial properties (such as, for example, increased nuclease resistance, increased uptake into cells, increased binding affinity for the RNA target) and a region that is a substrate for RNase H cleavage. In one preferred embodiment, a chimeric oligonucleotide comprises at least one region modified to increase target binding affinity, and, usually, a region that acts as a substrate for RNAse H. Affinity of an oligonucleotide for its target (in this case a nucleic acid encoding raf) is routinely determined by measuring the Tm of an oligonucleotide/target pair, which is the temperature at which the oligonucleotide and target dissociate; dissociation is detected spectrophotometrically. The higher the Tm, the greater the affinity of the oligonucleotide for the target. In a more preferred embodiment, the region of the oligonucleotide which is modified to increase raf mRNA binding affinity comprises at least one nucleotide modified at the 2' position of the sugar, most preferably a 2'-O-alkyl or 2'-fluoro-modified nucleotide. Such modifications are routinely incorporated into oligonucleotides and these oligonucleotides have been shown to have a higher Tm (i.e., higher target binding affinity) than 2'-deoxyoligonucleotides against a given target. The effect of such increased affinity is to greatly enhance antisense oligonucleotide inhibition of raf gene expression. RNAse H is a cellular endonuclease that cleaves the RNA strand of RNA:DNA duplexes; activation of this enzyme therefore results in cleavage of the RNA target, and thus can greatly enhance the efficiency of antisense inhibition. Cleavage of the RNA target can be routinely demonstrated by gel electrophoresis. In another preferred embodiment, the chimeric oligonucleotide is also modified to enhance nuclease resistance. Cells contain a variety of exo- and endo-nucleases which can degrade nucleic acids. A number of nucleotide and

nucleoside modifications have been shown to make the oligonucleotide into which they are incorporated more resistant to nuclease digestion than the native oligodeoxynucleotide. Nuclease resistance is routinely measured by incubating oligonucleotides with cellular extracts or isolated nuclease solutions and measuring the extent of intact oligonucleotide remaining over time, usually by gel electrophoresis. Oligonucleotides which have been modified to enhance their nuclease resistance survive intact for a longer time than unmodified oligonucleotides. A variety of oligonucleotide modifications have been demonstrated to enhance or confer nuclease resistance. Oligonucleotides which contain at least one phosphorothioate modification are presently more preferred. In some cases, oligonucleotide modifications which enhance target binding affinity are also, independently, able to enhance nuclease resistance.

Specific examples of some preferred oligonucleotides envisioned for this invention may contain phosphorothioates, phosphotriesters, methyl phosphonates, short chain alkyl or cycloalkyl intersugar linkages or short chain heteroatomic or heterocyclic intersugar ("backbone") linkages. Most preferred are phosphorothioates and those with CH_2-NH-O-CH_2, CH_2-N(CH_3)-O-CH_2 (known as the methylene(methylimino) or MMI backbone), CH_2-O-N(CH_3)-CH_2, CH_2-N(CH_3)-N(CH_3)-CH_2 and O-N(CH_3)-CH_2-CH_2 backbones (where phosphodiester is O-P-O-CH_2). Also preferred are oligonucleotides having morpholino backbone structures. Summerton, J.E. and Weller, D.D., U.S. Patent No: 5,034,506. In other preferred embodiments, such as the protein-nucleic acid or peptide-nucleic acid (PNA) backbone, the phosphodiester backbone of the oligonucleotide may be replaced with a polyamide backbone, the bases being bound directly or indirectly to the aza nitrogen atoms of the polyamide backbone. P.E. Nielsen, M. Egholm, R.H. Berg, O. Buchardt, *Science* 1991. 254, 1497. Other preferred oligonucleotides may contain alkyl and halogen-substituted sugar moieties comprising one of the following at the 2' position: OH, SH, SCH_3, F, OCN, OCH_3OCH_3, $OCH_3O(CH_2)_nCH_3$, $O(CH_2)_nNH_2$ or $O(CH_2)_nCH_3$ where n is from 1 to about

to C_{10} lower alkyl, substituted lower alkyl, alkaryl or aralkyl; Cl; Br; CN; CF_3; OCF_3; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; $SOCH_3$; SO_2CH_3; ONO_2; NO_2; N_3; NH_2; heterocycloalkyl; heterocycloalkaryl; aminoalkylamino; polyalkylamino; substituted silyl; an RNA cleaving group; a cholesteryl group; a conjugate; a reporter group; an intercalator; a group for improving the pharmacokinetic properties of an oligonucleotide; or a group for improving the pharmacodynamic properties of an oligonucleotide and other substituents having similar properties. Oligonucleotides may also have sugar mimetics such as cyclobutyls in place of the pentofuranosyl group. Other preferred embodiments may include at least one modified base form or "universal base" such as inosine.

The oligonucleotides in accordance with this invention preferably are from about 8 to about 50 nucleotides in length. In the context of this invention it is understood that this encompasses non-naturally occurring oligomers as hereinbefore described, having 8 to 50 monomers.

The oligonucleotides used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis. Equipment for such synthesis is sold by several vendors including Applied Biosystems. Any other means for such synthesis may also be employed; the actual synthesis of the oligonucleotides is well within the talents of the routineer. It is also well known to use similar techniques to prepare other oligonucleotides such as the phosphorothioates and alkylated derivatives. It is also well known to use similar techniques and commercially available modified amidites and controlled-pore glass (CPG) products such as biotin, fluorescein, acridine or psoralen-modified amidites and/or CPG (available from Glen Research, Sterling VA) to synthesize fluorescently labeled, biotinylated or other modified oligonucleotides such as cholesterol-modified oligonucleotides.

It has now been found that certain oligonucleotides targeted to portions of the c-raf mRNA are particularly useful for inhibiting raf expression and for interfering with cell

hyperproliferation. Methods for inhibiting c-raf expression using antisense oligonucleotides are, likewise, useful for interfering with cell hyperproliferation. In the methods of the invention, tissues or cells are contacted with oligonucleotides. In the context of this invention, to "contact" tissues or cells with an oligonucleotide or oligonucleotides means to add the oligonucleotide(s), usually in a liquid carrier, to a cell suspension or tissue sample, either *in vitro* or *ex vivo*, or to administer the oligonucleotide(s) to cells or tissues within an animal.

For therapeutics, methods of inhibiting hyperproliferation of cells and methods of treating abnormal proliferative conditions are provided. The formulation of therapeutic compositions and their subsequent administration is believed to be within the skill in the art. In general, for therapeutics, a patient suspected of needing such therapy is given an oligonucleotide in accordance with the invention, commonly in a pharmaceutically acceptable carrier, in amounts and for periods which will vary depending upon the nature of the particular disease, its severity and the patient's overall condition. The pharmaceutical compositions of this invention may be administered in a number of ways depending upon whether local or systemic treatment is desired, and upon the area to be treated. Administration may be topical (including ophthalmic, vaginal, rectal, intranasal), oral, or parenteral, for example by intravenous drip, intravenous injection or subcutaneous, intraperitoneal or intramuscular injection.

Formulations for topical administration may include ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms, gloves and the like may also be useful.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be

desirable.

Formulations for parenteral administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives.

In addition to such pharmaceutical carriers, cationic lipids may be included in the formulation to facilitate oligonucleotide uptake. One such composition shown to facilitate uptake is Lipofectin (BRL, Bethesda MD).

Dosing is dependent on severity and responsiveness of the condition to be treated, with course of treatment lasting from several days to several months or until a cure is effected or a diminution of disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body. Persons of ordinary skill can easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual oligonucleotides, and can generally be calculated based on EC50's in *in vitro* and *in vivo* animal studies. For example, given the molecular weight of compound (derived from oligonucleotide sequence and chemical structure) and an effective dose such as an IC50, for example (derived experimentally), a dose in mg/kg is routinely calculated.

The present invention is also suitable for diagnosing abnormal proliferative states in tissue or other samples from patients suspected of having a hyperproliferative disease such as cancer, psoriasis or blood vessel restenosis or atherosclerosis. The ability of the oligonucleotides of the present invention to inhibit cell proliferation may be employed to diagnose such states. A number of assays may be formulated employing the present invention, which assays will commonly comprise contacting a tissue sample with an oligonucleotide of the invention under conditions selected to permit detection and, usually, quantitation of such inhibition. Similarly, the present invention can be used to distinguish raf-associated tumors from tumors having other etiologies, in order that an efficacious treatment regimen can be designed.

The oligonucleotides of this invention may also be

used for research purposes. Thus, the specific hybridization exhibited by the oligonucleotides may be used for assays, purifications, cellular products preparations and in other methodologies which may be appreciated by persons of ordinary skill in the art.

The oligonucleotides of the invention are also useful for detection and diagnosis of raf expression. For example, radiolabeled oligonucleotides can be prepared by ^{32}P labeling at the 5' end with polynucleotide kinase. Sambrook et al., *Molecular Cloning. A Laboratory Manual*, Cold Spring Harbor Laboratory Press, 1989, Volume 3, p. 10.59. Radiolabeled oligonucleotides are then contacted with tissue or cell samples suspected of raf expression and the sample is washed to remove unbound oligonucleotide. Radioactivity remaining in the sample indicates bound oligonucleotide (which in turn indicates the presence of raf) and can be quantitated using a scintillation counter or other routine means. Radiolabeled oligo can also be used to perform autoradiography of tissues to determine the localization, distribution and quantitation of raf expression for research, diagnostic or therapeutic purposes. In such studies, tissue sections are treated with radiolabeled oligonucleotide and washed as described above, then exposed to photographic emulsion according to routine autoradiography procedures. The emulsion, when developed, yields an image of silver grains over the regions expressing raf. Quantitation of the silver grains permits raf expression to be detected.

Analogous assays for fluorescent detection of raf expression can be developed using oligonucleotides of the invention which are conjugated with fluorescein or other fluorescent tag instead of radiolabeling. Such conjugations are routinely accomplished during solid phase synthesis using fluorescently labeled amidites or CPG (e.g., fluorescein-labeled amidites and CPG available from Glen Research, Sterling VA. See 1993 Catalog of Products for DNA Research, Glen Research, Sterling VA, p. 21).

Each of these assays for raf is known in the art. One of skill could easily adapt these known assays for detection of

raf expression in accordance with the teachings of the invention providing a novel and useful means to detect raf expression.

Oligonucleotide inhibition of c-raf expression

The oligonucleotides shown in Table 1 were designed using the Genbank c-raf sequence HUMRAFR (Genbank listing x03484), synthesized and tested for inhibition of c-raf mRNA expression in T24 bladder carcinoma cells using a Northern blot assay. All are oligodeoxynucleotides with phosphorothioate backbones.

Table 1

Human c-raf Kinase Antisense Oligonucleotides

Isis #	Sequence (5' → 3')	Site	SEQ ID NO:
5000	TGAAGGTGAGCTGGAGCCAT	Coding	1
5074	GCTCCATTGATGCAGCTTAA	AUG	2
5075	CCCTGTATGTGCTCCATTGA	AUG	3
5076	GGTGCAAAGTCAACTAGAAG	STOP	4
5097	ATTCTTAAACCTGAGGGAGC	5'UTR	5
5098	GATGCAGCTTAAACAATTCT	5'UTR	6
5131	CAGCACTGCAAATGGCTTCC	3'UTR	7
5132	TCCCGCCTGTGACATGCATT	3'UTR	8
5133	GCCGAGTGCCTTGCCTGGAA	3'UTR	9
5148	AGAGATGCAGCTGGAGCCAT	Coding	10
5149	AGGTGAAGGCCTGGAGCCAT	Coding	11
6721	GTCTGGCGCTGCACCACTCT	3'UTR	12
6722	CTGATTTCCAAAATCCCATG	3'UTR	13
6731	CTGGGCTGTTTGGTGCCTTA	3'UTR	14
6723	TCAGGGCTGGACTGCCTGCT	3'UTR	15
7825	GGTGAGGGAGCGGGAGGCGG	5'UTR	16
7826	CGCTCCTCCTCCCCGCGGCG	5'UTR	17
7827	TTCGGCGGCAGCTTCTCGCC	5'UTR	18
7828	GCCGCCCCAACGTCCTGTCG	5'UTR	19
7848	TCCTCCTCCCCGCGGCGGGT	5'UTR	20
7849	CTCGCCCGCTCCTCCTCCCC	5'UTR	21
7847	CTGGCTTCTCCTCCTCCCCT	3'UTR	22
8034	CGGGAGGCGGTCACATTCGG	5'UTR	23
8094	TCTGGCGCTGCACCACTCTC	3'UTR	24

In a first round screen of oligonucleotides at concentrations of 100 nM or 200 nM, oligonucleotides 5074, 5075, 5132, 8034, 7826, 7827 and 7828 showed at least 50% inhibition of c-raf mRNA and these oligonucleotides are

these were preferred. Oligonucleotides 5132 and 7826 (SEQ ID NO: and SEQ ID NO: 17) showed greater than about 90% inhibition and are more preferred. In additional assays, oligonucleotides 6721, 7848, 7849 and 8094 decreased c-raf mRNA by greater than 50%. These oligonucleotides are also preferred. Of these, 7847 (SEQ ID NO: 22) showed greater than about 90% inhibition of c-raf mRNA and is more preferred.

Specificity of ISIS 5132 for ra

Specificity of ISIS 5132 for raf mRNA was demonstrated by a Northern blot assay in which this oligonucleotide was tested for the ability to inhibit Ha-ras mRNA as well as c-raf mRNA in T24 cells. Ha-ras is a cellular oncogene which is implicated in transformation and tumorigenesis. ISIS 5132 was shown to abolish c-raf mRNA almost completely with no effect on Ha-ras mRNA levels.

2'-modified oligonucleotides

Certain of these oligonucleotides were synthesized with either phosphodiester (P=O) or phosphorothioate (P=S) backbones and were also uniformly substituted at the 2' position of the sugar with either a 2'-O-methyl, 2'-O-propyl, or 2'-fluoro group. Oligonucleotides are shown in Table 2.

Table 2

...Uniformly 2' Sugar-modifie... c-raf Oligonucleotides

ISIS #	Sequence	Site	Modif.	SEQ ID NO:
6717	TCCCGCCTGTGACATGCATT	3'UTR	OMe/P=S	8
8033	CGGGAGGCGGTCACATTCGG	5'UTR	OMe/P=S	23
7829	GGTGAGGGAGCGGGAGGCGG	5'UTR	OMe/P=S	16
7830	CGCTCCTCCTCCCCGCGGCG	5'UTR	OMe/P=S	17
7831	TTCGGCGGCAGCTTCTCGCC	5'UTR	OMe/P=S	18
7832	GCCGCCCCAACGTCCTGTCG	5'UTR	OMe/P=S	19
7833	ATTCTTAAACCTGAGGGAGC	5'UTR	OMe/P=S	5
7834	GATGCAGCTTAAACAATTCT	5'UTR	OMe/P=S	6
7835	GCTCCATTGATGCAGCTTAA	AUG	OMe/P=S	2
7836	CCCTGTATGTGCTCCATTGA	AUG	OMe/P=S	3
8035	CGGGAGGCGGTCACATTCGG	5'UTR	OPr/P=O	23
7837	GGTGAGGGAGCGGGAGGCGG	5'UTR	OPr/P=O	16
7838	CGCTCCTCCTCCCCGCGGCG	5'UTR	OPr/P=O	17
7839	TTCGGCGGCAGCTTCTCGCC	5'UTR	OPr/P=O	18
7840	GCCGCCCCAACGTCCTGTCG	5'UTR	OPr/P=O	19
7841	ATTCTTAAACCTGAGGGAGC	5'UTR	OPr/P=O	5
7842	GATGCAGCTTAAACAATTCT	5'UTR	OPr/P=O	6
7843	GCTCCATTGATGCAGCTTAA	AUG	OPr/P=O	2
7844	CCCTGTATGTGCTCCATTGA	AUG	OPr/P=O	3
9355	CGGGAGGCGGTCACATTCGG	5'UTR	2'F/P=S	23

Oligonucleotides from Table 2 having uniform 2'O-methyl modifications and a phosphorothioate backbone were tested for ability to inhibit c-raf protein expression in T24 cells as determined by Western blot assay. Oligonucleotides 8033, 7834 and 7835 showed the greatest inhibition and are preferred. Of these, 8033 and 7834 are more preferred.

Chimeric oligonucleotides

Chimeric oligonucleotides having SEQ ID NO: 8 were prepared. These oligonucleotides had central "gap" regions of 6, 8, or 10 deoxynucleotides flanked by two regions of 2'-O-methyl modified nucleotides. Backbones were uniformly phosphorothioate. In Northern blot analysis, all three of these oligonucleotides (ISIS 6[illegible], 6-deoxy gap; ISIS 6717, 8-deoxy gap; ISIS 6729, 10-deoxy gap) showed greater than 70% inhibition of c-raf mRNA expression in T24 cells. These oligonucleotides are preferred. The 8-deoxy gap compound (6717) showed greater than 90% inhibition and is more preferred.

Additional chimeric oligonucleotides were synthesized having one or more regions of 2'-O-methyl modification and [illegible] phosphorothioate backbone. These are shown in Table 3. All are phosphorothioates; bold regions indicate 2'-O-methyl modified regions.

Table 3

Chimeric 2'-O-methyl P=S c-raf oligonucleotides

Isis #	Sequence	Target site	SEQ ID NO:
7848	TCCTCCTCCCCGCGGCGGGT	5'UTR	20
7852	**TCCTCCTCCCCGCGGCGGGT**	5'UTR	20
7849	CTCGCCCGCTCCTCCTCCCC	5'UTR	21
7851	CTCGCCCG**CTCCTCCTCCCC**	5'UTR	21
7856	TTCTCGCCCGCTCCTCCTCC	5'UTR	25
7855	**TTCTCGCCCGCTCCTCCTCC**	5'UTR	25
7854	**TTCTCCTCCTCCCCTGGCAG**	3'UTR	26
7847	CTGGCTTCTCCTCCTCCCCT	3'UTR	22
7850	CTGGC**TTCTCCTCCTCCCCT**	3'UTR	22
7853	CCTGCTGGC**TTCTCCTCCTC**	3'UTR	27

When tested for their ability to inhibit c-raf mRNA by Northern blot analysis, ISIS 7848, 7849, 7851, 7856, 7855, 7854, 7847, and 7853 gave better than 70% inhibition and are therefore preferred. Of these, 7851, 7855, 7847 and 7853 gave greater than 90% inhibition and are more preferred.

Additional chimeric oligonucleotides with various 2' modifications were prepared and tested. These are shown in Table 4. All are phosphorothioates; bold regions indicate 2'-modified regions.

Table 4

Chimeric 2'-modified P=S c-raf oligonucleotides

Isis #	Sequence	Target site	Modif.	SEQ ID
6720	TCCCGCCTGTGACATGCATT	3'UTR	2'-O-Me	8
6717	TCCCGCCTGTGACATGCATT	3'UTR	2'-O-Me	8
6729	TCCCGCCTGTGACATGCATT	3'UTR	2'-O-Me	8
8097	TCTGGCG**CTGCACCACTCTC**	3'UTR	2'-O-Me	24
9270	**TCCCGCCTGTGACATGCATT**	3'UTR	2'-O-Pro	8
9058	TCCCGCCTGTGACATGCATT	3'UTR	2'-F	8
9057	TCTGGC[illegible]	3'UTR	2'-F	24

Of these, oligonucleotides 6720, 6717, 6729, 9720 and 9058 are preferred. Oligonucleotides 6717, 6729, 9720 and 9058 are most preferred.

Two chimeric oligonucleotides with 2'-O-propyl sugar modifications and chimeric phosphorothioate backbones were also synthesized. These are shown in Table 5, in which italic regions indicate regions which are both 2'-modified and have phosphodiester backbones.

Table 5

Chimeric 2'-modified P=S/P=O c-raf oligonucleotides

Isis #	Sequence	Target site	Modif.	SEQ ID
9271	*TCCCGCCTGTGACATGCATT*	3'UTR	2'-O-Pro	8
8096	TCTGGCG*CTGCACCACTCTC*	3'UTR	2'-O-Pro	24

Inhibition of cancer cell proliferation

The phosphorothioate oligonucleotide ISIS 5132 was shown to inhibit T24 bladder cancer cell proliferation. Cells were treated with various concentrations of oligonucleotide in conjunction with lipofectin (cationic lipid which increases uptake of oligonucleotide). A dose-dependent inhibition of cell proliferation was demonstrated, as indicated in Table 6, in which "None" indicates untreated control (no oligonucleotide) and "Control" indicates treatment with negative control oligonucleotide. Results are shown as percent inhibition compared to untreated control.

Table 6

Inhibition of T24 Cell Proliferation by ISIS 5132

Oligo conc.	None	Control	5132
50 nM	0	[illegible]	23%
100 nM	0	[illegible]	24%
250 nM	0	[illegible]	74%
500 nM	0	[illegible]	82%

Effect of ISIS 5132 on T24 human bladder carcinoma tumors

Subcutaneous human T24 bladder carcinoma xenografts in nude mice were established and treated with ISIS 5132 and an unrelated control phosphorothioate oligonucleotide administered intraperitoneally three times weekly at a dosage of 25 mg/kg. In this preliminary study, ISIS 5132 inhibited tumor growth after eleven days by 31% compared to controls. Oligonucleotide-treated tumors remained smaller than control tumors throughout the course of the study.

Effect of ISIS 5132 on MDA-MB 231 human breast carcinoma tumors

Subcutaneous human MDA-MB 231 breast carcinoma xenografts in nude mice were established and treated with ISIS 5132 and an unrelated control phosphorothioate oligonucleotide administered intravenously once per day at a dosage of 0.6 mg/kg or 6.0 mg/kg. ISIS 5132 inhibited tumor growth after 27 days (end of study) by approximately 80% compared to controls.

ISIS 5132 was also effective when administered intraperitoneally to MDA-MB 231 xenografts in nude mice. Oligonucleotide was administered once per day at 0.6 mg/kg or 6.0 mg/kg. By day 27 (end of study), tumor volume was inhibited by 57% (0.6 mg/kg dose) or 64% (6.0 mg/kg) compared to control.

Effect of ISIS 5132 on c-raf RNA levels in MDA-MB231 tumors

RNA was isolated from MDA-MD231 tumor xenografts and Northern blots were performed to evaluate oligonucleotide effects on raf RNA levels. ISIS 5132 decreased raf RNA levels after 27 days by 67% when given intravenously and 39% when given intraperitoneally (both at 6 mg/kg).

Effect of ISIS 5132 on Colo 205 human colon carcinoma tumors

Subcutaneous human Colo 205 colon carcinoma xenografts in nude mice were established and treated with ISIS 5132 and an unrelated control phosphorothioate oligonucleotide administered intravenously once per day at a dosage of 6.0 mg/kg. In this study, ISIS 5132 inhibited tumor growth after 25 days by over

40% compared to controls.

[illegible] of ISIS 5132 on A549 human lung adenocarcinoma tumors

[illegible] subcutaneous human A549 lung adenocarcinoma xenografts were established in male Balb/c nude mice and treated with ISIS 5132 and a control oligonucleotide administered daily by intravenous injection at doses ranging from 0.006 to 6.0 mg/kg. ISIS 5132 decreased tumor size at all doses in a dose-dependent manner, as shown in Figure 1A. A scrambled raf oligonucleotide, ISIS 10353, had no effect at any dose (Figure 1B).

Effect of ISIS 5132 on c-raf RNA levels in A549 tumor cells

RNA was isolated from A549 tumor xenografts and Northern blots were performed to evaluate oligonucleotide effects on raf RNA levels. ISIS 5132 progressively decreased raf RNA levels beginning 8 hours after start of oligo treatment. When the experiment was terminated at day 13, RNA levels were still declining and had reached levels approximately 15% of control.

Effect of ISIS 6717, a 2'-O-methyl gapped oligonucleotide, on A549 lung xenograft tumors

ISIS 6717, a 2'-O-methyl gapped oligonucleotide shown in Table 4, was compared to ISIS 5132 for ability to inhibit A549 tumor xenograft growth. At doses of 0.006, 0.06, 0.6 and 6.0 mg/kg given intravenously, the two oligonucleotides were virtually indistinguishable in their effects on tumor growth. ISIS 6717 is therefore a preferred embodiment of this invention.

Antisense oligonucleotides targeted to A-raf

It is believed that certain oligonucleotides targeted to portions of the A-raf mRNA and which inhibit A-raf expression will be useful for interfering with cell hyperproliferation. Methods for inhibiting A-raf expression using such antisense oligonucleotides are [illegible] believed to be useful for interfering with cell hyperproliferation.

The phosphorothioate deoxyoligonucleotides shown in Table

7 were designed and synthesized using the Genbank A-raf sequence 'HUMARAFIR (Genbank listing x04790).

Table

Oligonucleotides Targeted to Human A-raf

Isis #	Sequence	Site	SEQ ID NO:
9060	GTC AAG ATG GGC TGA GGT GG	5' UTR	28
9061	CCA TCC CGG ACA GTC ACC AC	Coding	29
9062	ATG AGC TCC TCG CCA TCC AG	Coding	30
9063	AAT GCT GGT GGA ACT TGT AG	Coding	31
9064	CCG GTA CCC CAG GTT CTT CA	Coding	32
9065	CTG GGC AGT CTG CCG GGC CA	Coding	33
9066	CAC CTC AGC TGC CAT CCA CA	Coding	34
9067	GAG ATT TTG CTG AGG TCC GG	Coding	35
9068	GCA CTC CGC TCA ATC TTG GG	Coding	36
9069	CTA AGG CAC AAG GCG GGC TG	Stop	37
9070	ACG AAC ATT GAT TGG CTG GT	3' UTR	38
9071	GTA TCC CCA AAG CCA AGA GG	3' UTR	39
10228	CAT CAG GGC AGA GAC GAA CA	3' UTR	40

Oligonucleotides ISIS 9061, ISIS 9069 and ISIS 10228 were evaluated by Northern blot analysis for their effects on A-raf mRNA levels in A549, T24 and NHDF cells. All three oligonucleotides decreased A-raf RNA levels in a dose-dependent manner in all three cell types, with inhibition of greater than 50% at a 500 nM dose in all cell types. The greatest inhibition (88%) was achieved with ISIS 9061 and 9069 in T24 cells. These three oligonucleotides (ISIS 9061, 9069 and 10228) are preferred, with ISIS 9069 and 9061 being more preferred.

Identification of oligonucleotides targeted to rat and mouse c-raf

Many conditions which are believed to be mediated by raf kinase are not amenable to study in humans. For example, tissue graft rejection is a condition which is likely to be ameliorated by interference with raf expression, but, clearly, this must be evaluated in animals rather than human transplant patients. Another such example is restenosis. These conditions can be tested in animal models, however, such as the rat and mouse models used here.

oligonucleotide sequences for inhibiting c-raf expression in rat and mouse cells were identified. Rat and mouse c-raf genes have regions of high homology; a series of oligonucleotides which target both rat and mouse c-raf mRNA sequences were designed and synthesized, using information gained from evaluation of oligonucleotides targeted to human c-raf. These oligonucleotides were screened for activity in mouse bEND cells and rat A-10 cells using Northern blot assays. The oligonucleotides (all phosphorothioates) are shown in Table 8:

TABLE 8

Oligonucleotides targeted to mouse and rat c-raf

ISIS #	Target site	Sequence	SEQ ID:
10705	Coding	GGAACATCTGGAATTTGGTC	41
10706	Coding	GATTCACTGTGACTTCGAAT	42
10707	3'UTR	GCTTCCATTTCCAGGGCAGG	43
10708	3'UTR	AAGAAGGCAATATGGCTTTA	44
10709	3'UTR	GTGGTGCCTGCTGACTCTTC	45
10710	3'UTR	CTGGTGGCCTAAGAACAGCT	46
10711	AUG	GTATGTGCTCCATTGATGCA	47
10712	AUG	TCCCTGTATGTGCTCCATTG	48
11060	5'UTR	ATACTTATACCTGAGGGAGC	49
11061	5'UTR	ATGCATTCTGCCCCCAAGGA	50
11062	3'UTR	GACTTGTATACCTCTGGAGC	51
11063	3'UTR	ACTGGCACTGCACCACTGTC	52
11064	3'UTR	AAGTTCTGTAGTACCAAAGC	53
11065	3'UTR	CTCCTGGAAACCAGATTCAG	54

Oligonucleotides ISIS 11061 and 10707 were found to inhibit c-raf RNA levels by greater than 90% in mouse bEND cells at a dose of 400 nM. These two oligonucleotides inhibited raf RNA levels virtually entirely in rat A-10 cells at a concentration of 200 nM. The IC50 for ISIS 10707 was found to be 170 nM in mouse bEND cells and 85 nM in rat A-10 cells. The IC50 for ISIS 11061 was determined to be 40 nM in mouse bEND cells and 30 nM in rat A-10 cells.

Effect of ISIS-11061 on endogenous c-raf mRNA expression in mice

Mice were injected intraperitoneally with ISIS 11061 (50 mg/kg), control oligonucleotide or saline control once daily for [illegible] days. Animals were sacrificed and organs were analyzed for c-raf mRNA expression by Northern blot analysis. ISIS 11061 was found to decrease levels of c-raf mRNA in liver by approximately 70%. Control oligonucleotides had no effects on c-raf expression. The effect of ISIS 11061 was specific for c-raf; A-raf and G3PDH RNA levels were unaffected by oligonucleotide treatment.

Antisense oligonucleotide to c-raf increases survival in murine heart allograft model

To determine the therapeutic effects of the c-raf antisense oligonucleotide ISIS 11061 in preventing allograft rejection, this oligonucleotide was tested for activity in a murine vascularized heterotopic heart transplant model. Hearts from C57BI10 mice were transplanted into the abdominal cavity of C3H mice as primary vascularized grafts essentially as described by Isobe et al., *Circulation* **1991**, 84, 1246-1255. Oligonucleotides were administered by continuous intravenous administration via a 7-day Alzet pump. The mean allograft survival time for untreated mice was 7.83 ± 0.75 days(7, 7, 8, 8, 8, 9 days). Allografts in mice treated for 7 days with 20 mg/kg or 40 mg/kg ISIS 11061 all survived at least 11 days (11,11,12 days for 20 mg/kg dose and >11, >11, >11 days for the 40 mg/kg dose).

In a pilot study conducted in rats, hearts from Lewis rats were transplanted into the abdominal cavity of ACI rats. Rats were dosed with ISIS 11061 at 20 mg/kg for 7 days via Alzet pump. The mean allograft survival time for untreated rats was 8.86± 0.69 days (8, 8, 9, 9, 9, 9, 10 days). In rats treated with oligonucleotide, the allograft survival time was 15.3 ± 1.15 days (14, 16, 16 days).

Effects of antisense oligonucleotides targeted to c-raf on smooth muscle cell proliferation

smooth muscle cell proliferation is a cause of blood vessel [illegible] for example in atherosclerosis and restenosis after [illegible]. Experiments were performed to determine the effect of ISIS 11061 on proliferation of rat smooth muscle cells. Cells in culture were grown with and without ISIS 11061 (plus lipofectin) and cell proliferation was measured 24 and 48 hours after stimulation with fetal calf serum. ISIS 11061 (500 nM) was found to inhibit serum-stimulated cell growth in a dose-dependent manner with a maximal inhibition of 46% and 75% at 24 hours and 48 hours, respectively. An IC50 value of 200 nM was obtained for this compound. An unrelated control oligonucleotide had no effect at doses up to 500 nM.

Effects of antisense oligonucleotides targeted to c-raf on restenosis in rats

A rat carotid artery injury model of angioplasty restenosis has been developed and has been used to evaluate the effects on restenosis of antisense oligonucleotides targeted to the c-myc oncogene. Bennett et al., *J. Clin. Invest.* 1994, 93, 820-828. This model will be used to evaluate the effects of antisense oligonucleotides targeted to rat c-raf, particularly ISIS 11061, on restenosis. Following carotid artery injury with a balloon catheter, oligonucleotides are administered either by intravenous injection, continuous intravenous administration via Alzet pump, or direct administration to the carotid artery in a pluronic gel matrix as described by Bennett et al. After recovery, rats are sacrificed, carotid arteries are examined by microscopy and effects of treatment on luminal cross-sections are determined.

The invention is further illustrated by the following examples which are illustrative only and are not intended to limit the present invention to specific embodiments.

EXAMPLES

Example 1 Synthesis and Characterization of Oligonucleotides

Unmodified DNA oligonucleotides were synthesized on an automated DNA synthesizer (Applied Biosystems model 380B) using standard phosphoramidite chemistry with oxidation by iodine. ß-cyanoethyldiisopropyl phosphoramidites were purchased from

Applied Biosystems (Foster City, CA). For phosphorothioate oligonucleotides the standard oxidation bottle was replaced by 0.2 M solution of 3H-1,2-benzodithiole-3-one 1,1-dioxide in acetonitrile for the stepwise thiation of the phosphite linkages. The thiation cycle wait step was increased to 68 seconds and was followed by the capping step. 2'-O-methyl phosphorothioate oligonucleotides were synthesized using 2'-O-methyl β-cyanoethyldiisopropyl-phosphoramidites (Chemgenes, Needham MA) and the standard cycle for unmodified oligonucleotides, except the wait step after pulse delivery of tetrazole and base was increased to 360 seconds. The 3'-base used to start the synthesis was a 2'-deoxyribonucleotide. 2'-O-propyl oligonucleotides were prepared by a slight modification of this procedure.

2'-fluoro phosphorothioate oligonucleotides were synthesized using 5'-dimethoxytrityl-3'-phosphoramidites and prepared as disclosed in U.S. patent application Serial No. 463,358, filed January 11, 1990, and 566,977, filed August 13, 1990, which are assigned to the same assignee as the instant application and which are incorporated by reference herein. The 2'-fluoro oligonucleotides were prepared using phosphoramidite chemistry and a slight modification of the standard DNA synthesis protocol: deprotection was effected using methanolic ammonia at room temperature.

After cleavage from the controlled pore glass column (Applied Biosystems) and deblocking in concentrated ammonium hydroxide at 55°C for 18 hours, the oligonucleotides were purified by precipitation twice out of 0.5 M NaCl with 2.5 volumes ethanol. Analytical gel electrophoresis was accomplished in 20% acrylamide, 8 M urea, 45 mM Tris-borate buffer, pH 7.0. Oligodeoxynucleotides and their phosphorothioate analogs were judged from electrophoresis to be greater than 80% full length material.

Example 2 Northern blot analysis of inhibition of c-raf mRNA expression

The human urinary bladder tumor cell line T24 was obtained from the American Type Culture Collection (Rockville MD). Cells

were grown in McCoy's 5A medium with L-glutamine (Gibco BRL, Gaithersburg, MD) supplemented with 10% heat-inactivated fetal bovine serum and 50 U/ml each of penicillin and streptomycin. Cells were seeded on 100 mm plates. When they reached 70% confluency, they were treated with oligonucleotide. Plates were washed with 10 ml prewarmed PBS and 5 ml of Opti-MEM reduced-serum medium containing 2.5 µl LTMA. Oligonucleotide with lipofectin was then added to the desired concentration. After 4 hours of treatment, the medium was replaced with McCoy's medium. Cells were harvested 24 to 72 hours after oligonucleotide treatment and RNA was isolated using a standard CsCl purification method. Kingston, R.E., in *Current Protocols in Molecular Biology*, (F.M. Ausubel, R. Brent, R.E. Kingston, D.D. Moore, J.A. Smith, J.G. Seidman and K. Strahl, eds.), John Wiley and Sons, NY. Total RNA was isolated by centrifugation of cell lysates over a CsCl cushion. RNA samples were electrophoresed through 1.2% agarose-formaldehyde gels and transferred to hybridization membranes by capillary diffusion over a 12-14 hour period. The RNA was cross-linked to the membrane by exposure to UV light in a Stratalinker (Stratagene, La Jolla, CA) and hybridized to random-primed ^{32}P-labeled c-raf cDNA probe (obtained from ATCC) or G3PDH probe as a control. RNA was quantitated using a Phosphorimager (Molecular Dynamics, Sunnyvale, CA).

Example 3 Specific inhibition of c-raf kinase protein expression in T24 cells

T24 cells were treated with oligonucleotide (200 nM) and lipofectin at T=0 and T=24 hours. Protein extracts were prepared at T=48 hours, electrophoresed on acrylamide gels and analyzed by Western blot using polyclonal antibodies against c-raf (UBI, Lake Placid, NY) or A-raf (Transduction Laboratories, Knoxville, TN). Radiolabeled secondary antibodies were used and raf protein was quantitated using a Phosphorimager (Molecular Dynamics, Sunnyvale CA).

Example 4 Antisense inhibition of cell proliferation

T24 cells were treated on day 0 for two hours with various concentrations of oligonucleotide and lipofectin (50 nM oligonucleotide in the presence of 2 µg/ml lipofectin; 100 nM oligonucleotide and 2µg/ml lipofectin; 250 nM oligonucleotide and 5 µg/ml lipofectin or 500 nM oligonucleotide and 10 µg/ml lipofectin). On day 1, cells were treated for a second time at desired oligonucleotide concentration for two hours. On day 2, cells were counted.

Example 5 Effect of ISIS 5132 on T24 Human Bladder Carcinoma Tumor Xenografts in Nude Mice

5×10^6 T24 cells were implanted subcutaneously in the right inner thigh of nude mice. Oligonucleotides (ISIS 5132 and an unrelated control phosphorothioate oligonucleotide suspended in saline) were administered three times weekly beginning on day 4 after tumor cell inoculation. A saline-only control was also given. Oligonucleotides were given by intraperitoneal injection. Oligonucleotide dosage was 25 mg/kg. Tumor size was measured and tumor volume was calculated on the eleventh, fifteenth and eighteenth treatment days.

Example 6 Effect of ISIS 5132 on MDA-MB 231 Human Breast Carcinoma Tumor Xenografts in Nude Mice

5×10^6 MDA-MB 231 cells were implanted subcutaneously in the right inner thigh of nude mice. Oligonucleotides (ISIS 5132 and an unrelated control phosphorothioate oligonucleotide suspended in saline) were administered once daily beginning on day 10 after tumor cell inoculation. A saline-only control was also given. Oligonucleotides were given by intravenous injection at a dosage of 0.6 mg/kg or 6.0 mg/kg. Tumor size was measured and tumor volume was calculated on days 10, 13, 16, 20, 23 and 27 following tumor cell inoculation.

For intraperitoneal oligonucleotide administration oligonucleotides were administered once daily beginning on day 10 after tumor cell inoculation. A saline-only control was also given. Oligonucleotides were given by intraperitoneal injection at a dosage of 0.6 mg/kg or 6.0 mg/kg. Tumor size was measured

and tumor volume was calculated on days 10, 13, 16, 20, 23 and 27 following tumor cell inoculation.

Example [illegible] Effect of ISIS 5132 on Colo 205 Human Colon Carcinoma Tumor Xenografts in [illegible] Mice

[illegible] x 10^6 Colo 205 cells were implanted subcutaneously in the right inner thigh of nude mice. Oligonucleotides (ISIS 5132 and an unrelated control phosphorothioate oligonucleotide suspended in saline) were administered once per day beginning on day 5 after tumor cell inoculation. A saline-only control was also given. Oligonucleotides were given by intravenous injection. Oligonucleotide dosage was 6 mg/kg. Tumor size was measured and tumor volume was calculated on days 5, 8, 11, 14, 18, 22 and 25 after tumor inoculation.

Example 8 Diagnostic Assay for raf-associated Tumors Using Xenografts in Nude Mice

Tumors arising from raf expression are diagnosed and distinguished from other tumors using this assay. A biopsy sample of the tumor is treated, e.g., with collagenase or trypsin or other standard methods, to dissociate the tumor mass. 5 x 10^6 tumor cells are implanted subcutaneously in the inner thighs of two or more nude mice. Antisense oligonucleotide (e.g., ISIS 5132) suspended in saline is administered to one or more mice by intraperitoneal injection three times weekly beginning on day 4 after tumor cell inoculation. Saline only is given to a control mouse. Oligonucleotide dosage is 25 mg/kg. Tumor size is measured and tumor volume is calculated on the eleventh treatment day. Tumor volume of the oligonucleotide-treated mice is compared to that of the control mouse. The volume of raf-associated tumors in the treated mice are measurably smaller than tumors in the control mouse. Tumors arising from causes other than raf expression are not expected to respond to the oligonucleotides targeted to raf and, therefore, the tumor volumes of oligonucleotide-treated and control mice are equivalent.

Example 9 Detection of raf expression

Oligonucleotides are radiolabeled after synthesis by ^{32}P labeling at the 5' end with polynucleotide kinase. Sambrook et al., *Molecular Cloning. A Laboratory Manual*, Cold Spring Harbor Laboratory Press, 1989, Volume 2, 11.31-11.32. Radiolabeled oligonucleotides are contacted with tissue or cell samples suspected of raf expression, such as tumor biopsy samples or skin samples where psoriasis is suspected, under conditions in which specific hybridization can occur, and the sample is washed to remove unbound oligonucleotide. Radioactivity remaining in the sample indicates bound oligonucleotide and is quantitated using a scintillation counter or other routine means.

Radiolabeled oligonucleotides of the invention are also used in autoradiography. Tissue sections are treated with radiolabeled oligonucleotide and washed as described above, then exposed to photographic emulsion according to standard autoradiography procedures. The emulsion, when developed, yields an image of silver grains over the regions expressing raf. The extent of raf expression is determined by quantitation of the silver grains.

Analogous assays for fluorescent detection of raf expression use oligonucleotides of the invention which are labeled with fluorescein or other fluorescent tags. Labeled DNA oligonucleotides are synthesized on an automated DNA synthesizer (Applied Biosystems model 380B) using standard phosphoramidite chemistry with oxidation by iodine. ß-cyanoethyldiisopropyl phosphoramidites are purchased from Applied Biosystems (Foster City, CA). Fluorescein-labeled amidites are purchased from Glen Research (Sterling VA). Incubation of oligonucleotide and biological sample is carried out as described for radiolabeled oligonucleotides except that instead of a scintillation counter, a fluorimeter or fluorescence microscope is used to detect the fluorescence which indicates raf expression.

Example 10: A549 x nograft

5 x 10^6 A549 cells were implanted subcutaneously in the inner thigh of nude mice. Oligonucleotides (ISIS 5132 and a

scrambled raf control phosphorothioate oligonucleotide, ISIS 10357) suspended in saline were administered once daily by intravenous injection at doses ranging from 0.006 to 6.0 mg/kg. Resulting tumors were measured on days 9, 12, 17 and 21 and tumor volumes were calculated.

Example 11: Effect of oligonucleotide on endogenous c-raf expression

Mice were treated by intraperitoneal injection at an oligonucleotide dose of 50 mg/kg on days 1, 2 and 3. On day 4 animals were sacrificed and organs removed for c-raf mRNA assay by Northern blot analysis. Four groups of animals were employed: 1) no oligonucleotide treatment (saline); 2) negative control oligonucleotide ISIS 1082 (targeted to herpes simplex virus); 3) negative control oligonucleotide 4189 (targeted to mouse protein kinase C-α); 4) ISIS 11061 targeted to rodent c-raf.

Example 12: Cardiac allograft rejection model

Hearts were transplanted into the abdominal cavity of rats or mice (of a different strain from the donor) as primary vascularized grafts essentially as described by Isobe et al., *Circulation* **1991**, 84, 1246-1255. Oligonucleotides were administered by continuous intravenous administration via a 7-day Alzet pump. Cardiac allograft survival was monitored by listening for the presence of a second heartbeat in the abdominal cavity.

Example 13: Proliferation assay using rat A-10 smooth muscle cells

A10 cells were plated into 96-well plates in Dulbecco's modified Eagle medium (DMEM) + 10% fetal calf serum and allowed to attach for 24 hours. Cells were made quiescent by the addition of DMEM + 0.2% dialyzed fetal calf serum for an additional 24 hours. During the last 4 hours of quiescence, cells were treated with ISIS 11061 + lipofectin (Gibco-BRL, Bethesda MD) in serum-free medium. Medium was then removed, replaced with fresh medium and the cells were stimulated with 10% fetal calf serum. The plates were the placed into the

[illegible] evaluated by MTS conversion ([illegible] Promega Cell Proliferation Kit) at 24 and 48 hours [illegible] of oligonucleotide, ISIS [illegible] ([illegible] herpes simplex virus) was also tested.

Example 14: Rat carotid artery restenosis model

This model has been described by Bennett et al., J. Clin. Invest. 1994, 93, 820-828. Intimal hyperplasia is induced by balloon catheter dilatation of the carotid artery of the rat. Rats are anesthetized and common carotid artery injury is induced by passage of a balloon embolectomy catheter distended with 20 µl of saline. Oligonucleotides are applied to the adventitial surface of the arterial wall in a pluronic gel solution. Oligonucleotides are dissolved in a 0.25% pluronic gel solution at 4°C (F127, BASF Corp.) at the desired dose. 100 µl of the gel solution is applied to the distal third of the common carotid artery immediately after injury. Control rats are treated similarly with gel containing control oligonucleotide or no oligonucleotide. The neck wounds are closed and the animals allowed to recover. 14 days later, rats are sacrificed, exsanguinated and the carotid arteries fixed in situ by perfusion with paraformaldehyde and glutaraldehyde, excised and processed for microscopy. Cross-sections of the arteries are calculated.

In an alternative to the pluronic gel administration procedure, rats are treated by intravenous injection or continuous intravenous infusion (via Alzet pump) of oligonucleotide.

SEQUENCE LISTING

(i) APPLICATION

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(iii) TITLE OF INVENTION: Antisense Oligonucleotide Modulation of raf Gene Expressi

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(E) COUNTRY: USA

(F) ZIP: 08002

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: DISKETTE, 3.5 INCH, 1.44 Mb STORAGE

(B) COMPUTER: IBM PS/2

(C) OPERATING SYSTEM: PC-DOS

(D) SOFTWARE: WORDPERFECT 5.1

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: N/A

(B) FILING DATE: Herewith

(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: 08/250,856

(B) FILING DATE: May 31, 1995

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME [illegible]

(B) REGISTRATION [illegible]

REFERENCE/DOCKET NUMBER: MSPH 0134

(1) [illegible] INFORMATION:

(A) ADDRESSEE: [illegible] 709, 779 [illegible] 0

(B) TELEFAX: (609) 779-8

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

TGAAGGTGAG CTGGAGCCAT (20)

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GCTCCATTGA TGCAGCTTAA (20)

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

GGTGCAAAGT CAACTAGAAG (20)

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

ATTCTTAAAC CTGAGGGAGC (20)

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE. Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

CAGCACTGCA AATGGCTTCC (20)

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

TCCCGCCTGT GACATGCATT (20)

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

AGAGATGCAG CTGGAGCCAT (20)

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

AGGTGAAGGC CTGGAGCCAT (20)

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

GTCTGCCAGC TTCCTTGGAA (20)

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

TCGATTCCTA AANTCCCAIG (20)

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

CTGGGCTGTT TGGTGCCTTA (20)

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Singl

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

[illegible] (20)

(2) INFORMATION FOR SEQ ID NO: [illegible]

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

CGCTCCTCCT CCCCGCGGCG (20)

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

CGCTCCTCCT CCCCGCGGCG (20)

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

TTCGGCGGCA GCTTCTCGCC (20)

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

GCCGCCCCAA CGTCCTGTCG (20)

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

TCCTCCTCCC CGCGGCGGGT (20)

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

CTCGCCCCCT CCTCCTCCCC (20)

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: [illegible]

[illegible]

[illegible]

[illegible]

[illegible]

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22.

AGGTTTCTC CTCCTCCTCT (20)

(2) INFORMATION FOR SEQ ID NO: 23

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

CGGGAGGCGG TCACATTCGG (20)

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24.

TCTGGCGCTG CACCACTCTC (20)

(2) INFORMATION FOR SEQ ID NO: 25

(i) SEQUENCE CHARACTERISTICS

[illegible]

[illegible]

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

TTCTCCTCCT CCCCTGGCAG (20)

(2) INFORMATION FOR SEQ ID NO: 27:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

CCTGCTGGCT TCTCCTGCTC (20)

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(ii) TOPOLOGY: Linear

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ix) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

GLATCCGGA CATTCACCAC (20)

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ix) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

ATGAACTCCT GGCCATCCAG (20)

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

[illegible] NO. 31:

[illegible] DNA

[illegible] NO. 32:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

CCGGTACCCC AGGTTCTTCA (20)

(2) INFORMATION FOR SEQ ID NO: 33:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

CTGGGCAGTC TGCCGGGCCA (20)

(2) INFORMATION FOR SEQ ID NO: 34:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

[illegible]

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

GAGATTTGC TGAGGTCCGG (20)

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO. 36:

GCACTCCCCT CAATCTTGGG (20)

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

ACGAACATTG ATTGGCTGGT (20)

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:

GTATCCTCAA AGCCAAGAGG (20)

(2) INFORMATION FOR SEQ ID NO: 40:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 40:

GATCAGGTCA GAGACAACCA (20)

(ii) INFORMATION FOR SEQ ID NO: 40:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 41:

GGAACATCTG GAATTGGTC (20)

(2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42 :

GATTCACTGT GACTTCGAAT (20)

(2) INFORMATION FOR SEQ ID NO: 43:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE. Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 43:

GCATCGATTT CGAGCGTGTG (20)

(2) INFORMATION FOR SEQ ID NO: 44

(i) [illegible]

(ii) [illegible]: Single

(iii) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

AAGAAGGCAA TATGAAGTTA (20)

(2) INFORMATION FOR SEQ ID NO: 45:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

GTGGTGCCTG CTGACTCTTC (20)

(2) INFORMATION FOR SEQ ID NO: 46:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

CTGGTGGCCT AAGAACAGCT (20)

(2) INFORMATION FOR SEQ ID NO: 47:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

GTGTCTCTTC CATTGATGCA (20)

(2) INFORMATION FOR SEQ ID NO: 48:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

TCCCTGTATG TGCTCCATTG (20)

(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

ATACTTATAC CTGAGGGAGC (20)

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS

(A) LENGTH: 20

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

GACTTGTATA CCTCTGGAGC (20)

(2) INFORMATION FOR SEQ ID NO: 52:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

ACTGGCACTG CACCACTGTC (20)

(2) INFORMATION FOR SEQ ID NO: 53:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

AACTTCTGTA GTACCAAAGC (20)

(2) INFORMATION FOR SEQ ID NO: 54:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: Nucleic Acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(iv) ANTI-SENSE: Yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

CTCCTGCAAG ACAGATTCAG (20)

What is Claimed is:

1. An oligonucleotide 8 to 40 nucleotides in length [illegible] which is capable of binding RNA encoding

2. The oligonucleotide of claim 1 which is targeted to RNA encoding human A-raf.

3. The oligonucleotide of claim 2 having SEQ ID NO 25, 31 or 40.

4. The oligonucleotide of claim 1 which is targeted to mRNA encoding human c-raf.

5. The oligonucleotide of claim 4 which is targeted to a translation initiation site, 3' untranslated region or 5' untranslated region of mRNA encoding human c-raf

6. The oligonucleotide of claim 1 which has at least one phosphorothioate linkage.

7. The oligonucleotide of claim 1 wherein at least one of the nucleotide units of the oligonucleotide is modified at the 2' position of the sugar moiety.

8. The oligonucleotide of claim 7 wherein said modification at the 2' position of the sugar moiety is a 2'-O-alkyl or a 2'-fluoro modification.

9. The oligonucleotide of claim 1 in a pharmaceutically acceptable carrier.

10. The oligonucleotide of claim 5 comprising SEQ ID NO: 2, 6, 8 12, 17, 20, 21, 22, 24, 25, 26 or 27.

11. An oligonucleotide having SEQ ID NO 9.

12. The oligonucleotide of claim 1 comprising at least [illegible] modification.

13. A chimeric oligonucleotide 8 to 30 nucleotides in length which is targeted to nucleic acid encoding human raf which comprises a first region having at least one nucleotide which is modified to enhance target affinity and a second region which is a substrate for RNAse H, said chimeric oligonucleotide being capable of inhibiting raf expression.

14. The oligonucleotide of claim 13 wherein the nucleotide which is modified to enhance target affinity is modified at the 2' position of the sugar moiety.

15. The oligonucleotide of claim 14 wherein the modification at the 2' position of the sugar moiety is a 2'-O-alkyl or a 2'-fluoro modification.

16. The oligonucleotide of claim 13 wherein the region which is a substrate for RNAse H compriaes at least one 2' deoxynucleotide.

17. The oligonucleotide of claim 13 which has at least one phosphorothioate linkage.

18. The oligonucleotide of claim 13 which is targeted to a translation initiation site, 3' untranslated region or 5' untranslated region of mRNA encoding human c-raf.

19. The oligonucleotide of claim 13 in a pharmaceutically acceptable carrier.

20. The oligonucleotide of claim 1 comprising SEQ ID NO: 8, 21, 24, 25, 26 or 27.

21. A chimeric oligonucleotide having SEQ ID NO: 8 which contains a first region having at least one nucleotide which is

modified to enhance target affinity and a second region which is a substrate for RNase H, said chimeric oligonucleotide being capable of inhibiting raf expression.

23. A method of inhibiting the expression of human raf comprising contacting tissues or cells which express human raf with an oligonucleotide 8 to 50 nucleotides in length which is complementary to mRNA encoding human raf and which is capable of inhibiting raf expression.

23. The method of claim 22 wherein said oligonucleotide is targeted to mRNA encoding human A raf.

24. The method of claim 22 wherein said oligonucleotide is targeted to mRNA encoding human c-raf.

25. The method of claim 24 wherein said oligonucleotide is targeted to a translation initiation site, 3' untranslated region or 5' untranslated region of mRNA encoding human c-raf.

26. The method of claim 25 wherein said oligonucleotide comprises SEQ ID NO: 2, 6, 8, 12, 17, 20, 21, 22, 23, 24, 25, 26 or 27.

27. The method of claim 22 wherein said expression of human raf is abnormal expression.

28. The method of claim 22 wherein said oligonucleotide is a chimeric oligonucleotide.

29. The method of claim 28 wherein said chimeric oligonucleotide comprises SEQ ID NO: 8, 21, 24, 25, 26 or 27.

30. The method of claim 22 wherein said oligonucleotide has at least one phosphorothioate linkage.

31. A method of inhibiting the expansion of hyperproliferating tissues or cells which expresses h-ras comprising contacting tissues or cells which express h-ras with an oligonucleotide having SEQ ID NO: 1.

32. A method of inhibiting hyperproliferation of cells comprising contacting hyperproliferating cells with an oligonucleotide 8 to 50 nucleotides in length which is targeted to mRNA encoding human raf and which is capable of inhibiting raf expression.

33. The method of claim 32 wherein said oligonucleotide is targeted to mRNA encoding human A-raf.

34. The method of claim 32 wherein said oligonucleotide is targeted to mRNA encoding human c-raf.

35. The method of claim 34 wherein said oligonucleotide is targeted to a translation initiation site, 3' untranslated region or 5' untranslated region of mRNA encoding human c-raf.

36. The method of claim 35 wherein said oligonucleotide comprises SEQ ID NO: 2, 6, 8, 12, 17, 20, 21, 22, 23, 24, 25, 26 or 27.

37. The method of claim 32 wherein said oligonucleotide is a chimeric oligonucleotide.

38. The method of claim 37 wherein said chimeric oligonucleotide comprises SEQ ID NO: 8, 21, 24, 25, 26 or 27.

39. The method of claim 32 wherein said oligonucleotide has at least one phosphorothioate linkage.

40. A method of inhibiting hyperproliferation of cells comprising contacting hyperproliferating cells with an oligonucleotide having SEQ ID NO: 3

41. A method of treating an animal proliferative condition comprising contacting a subject or cell in said animal or body fluid of said subject suspected of having an abnormal proliferative condition with an oligonucleotide 8 to 30 nucleotides in length which is targeted to mRNA encoding human raf and which is capable of inhibiting raf expression.

42. The method of claim 41 wherein the condition is hyperproliferative disorder.

43. The method of claim 42 wherein the hyperproliferative disorder is cancer, restenosis, psoriasis or a disorder characterized by T-cell activation and growth.

44. The method of claim 41 wherein said oligonucleotide is targeted to mRNA encoding human A-raf

45. The method of claim 41 wherein said oligonucleotide is targeted to mRNA encoding human c-raf

46. The method of claim 45 wherein said oligonucleotide is targeted to the 3' untranslated region or the 5' untranslated region of mRNA encoding human c-raf.

47. The method of claim 46 wherein said oligonucleotide comprises SEQ ID NO: 2, 6, 8, 12, 17, 20, 21, 22, 23, 24, 25 or 27.

48. The method of claim 41 wherein said oligonucleotide is a chimeric oligonucleotide

49. The method of claim 48 wherein said chimeric oligonucleotide comprises SEQ ID NO: 8, 21, 24, 25, 26 or 27.

50. The method of claim 41 wherein said oligonucleotide has at least one phosphorothioate linkage.

51. A method of treating an abnormal proliferative
condition comprising contacting a subject, or a bodily
fluid of said subject, suspected of having an abnormal
proliferative condition with an oligonucleotide having SEQ
ID NO:3.

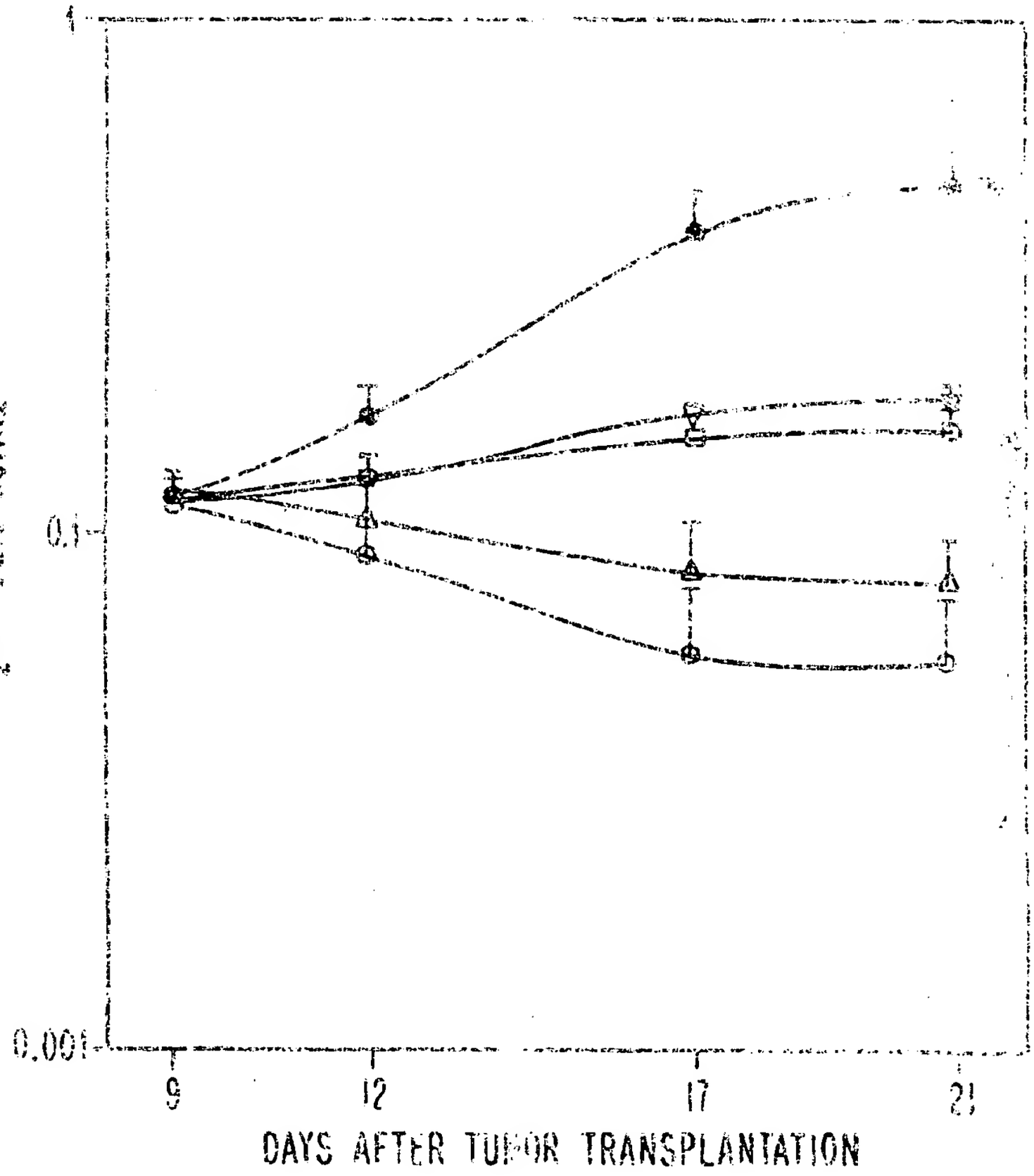
CONTROLS
0.10 mg/kg i.v.
0.30 mg/kg i.v.
0.96 mg/kg i.v.
TUMOR SIZE, cm³
1.0
0.1
0.001
9
12
17
21
DAYS AFTER TUMOR TRANSPLANTATION

Fig. 1A

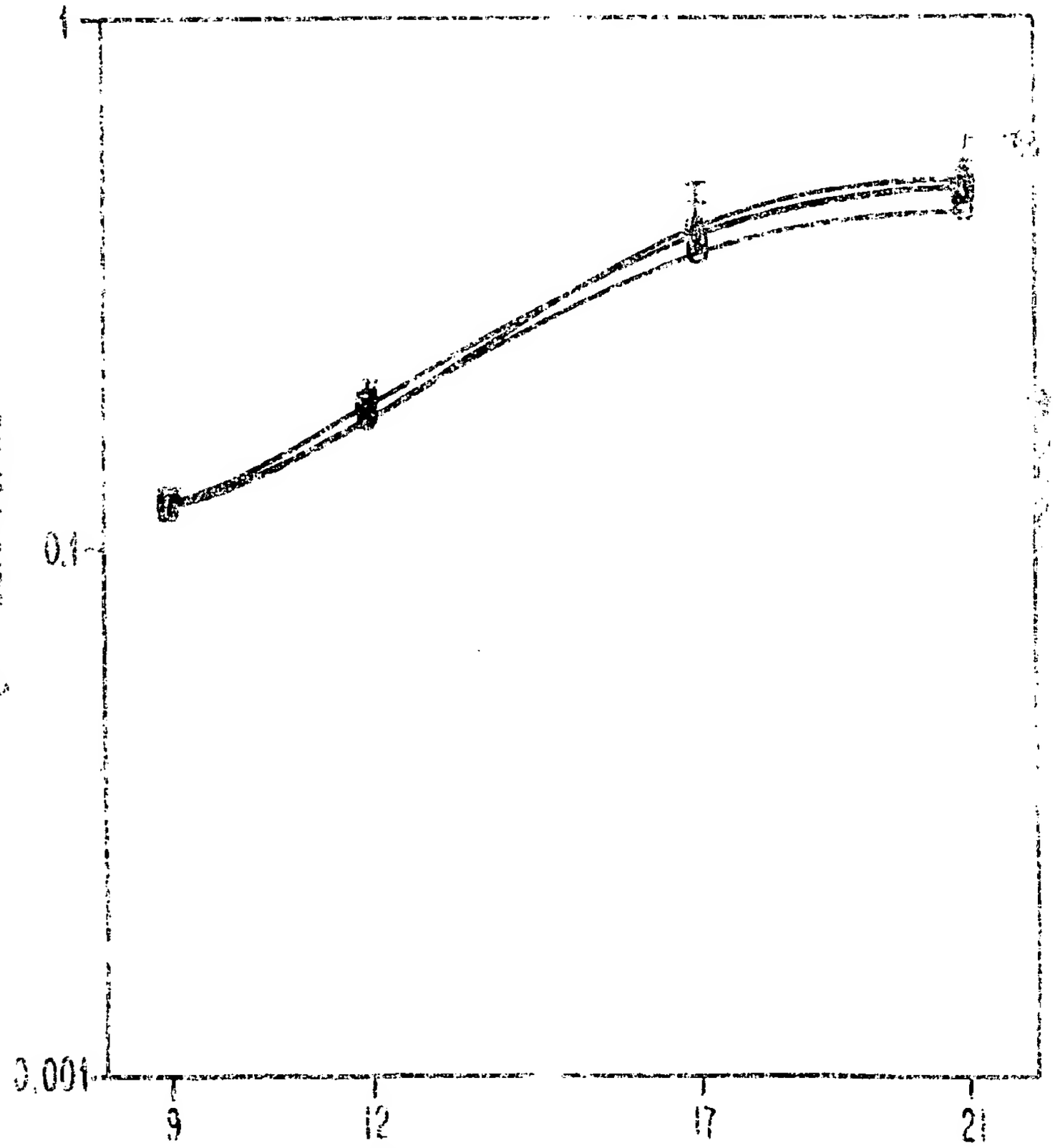

Fig. 1B

[illegible]

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
[illegible]	WO, A, 94/15645 (TEXAS BIOTECHNOLOGY) [illegible] July 1994, see entire document.	[illegible] 2 [illegible] 41, 43, [illegible] 48 [illegible]
[illegible]		3, 7, 8, [illegible] 38, [illegible] 43, [illegible] 49, 50
Y, P	WO, A, 94/23755 (BOARD OF REGENTS OF THE UNIVERSITY OF NEBRASKA) 27 October 1994, see entire document.	1-51

[illegible] documents are listed in the continuation of Box C. See patent family annex.

*	Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A"	document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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Date of the actual completion of the international search [illegible] 1995	Date of mailing of the international search report [illegible] SEP 1995
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231	Authorized officer Thomas G. Larson, Ph.D.
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INTERNATIONAL SEARCH REPORT [illegible]

International application No. [illegible]

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
[illegible]	US, A, [illegible] [illegible], see entire document.	1-[illegible]
[illegible]	WO, A, 9[illegible] (THE UNITED STATES OF AMERICA AS REPRESENTED BY THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES) [illegible], see entire document.	[illegible]
X — Y	European Journal of Immunology, Vol. 2[illegible], issued 199[illegible], Riedel et al, "The Mitogenic Response of T Cells to Interleukin-2 Requires Raf-1," pages 31[illegible]-3150, see entire document.	1-[illegible], 9, [illegible] 24, [illegible], 33 — [illegible]
Y	Journal of Biological Chemistry, Vol. [illegible], No. [illegible], issued [illegible], [illegible] et al, "Inhibition of Raf-1 [illegible] Expression [illegible] Insulin-Stimulated [illegible] Synthesis in [illegible] Cells," pages [illegible], see entire document.	[illegible]
Y	Science, Vol. 243, issued 10 March 1989, Kasid et al, "Effect of Antisense c-raf-1 on Tumorigenicity and Radiation Sensitivity of a Human Squamous Carcinoma," pages 1354-1356, see entire document.	[illegible]
Y	Nucleic Acids Research, Vol. 15, No. 2, issued 1987, Beck et al, "The Complete Coding Sequence of the Human A-raf-1 Oncogene and Transforming Activity of a Human A-raf Carrying Retrovirus," pages 595-609, see entire document.	1-3, 6-9, 12-19, 22-23, 27, 29-32, 37, 39, 41-44, 48-50
Y	Nucleic Acids Research, Vol. 14, No. 2, issued 1986, Bonner et al, "The Complete Coding Sequence of the Human raf Oncogene and the Corresponding Sequence of the c-raf-1 Gene," pages 1009-1015, see entire document.	1, 6-22, 24-32, 34-45, 47-5[illegible]
Y	Critical Reviews in Oncogenesis, Vol. 3, Nos. 1 and 2, issued 1992, Neckers et al, "Antisense Inhibition of Oncogene Expression," pages 175-230, see pages 177-179, 181-183, 186-187, 190-196, 202-213.	1-51
Y	Chemical Reviews, Vol. 90, No. 4, issued June 1990, Uhlmann et al, "Antisense Oligonucleotides: A New Therapeutic Principle," pages 544-584, see pages 548-553, 558, 571-573.	6-9, 12-21, [illegible]-30, 37-39, 48-53

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US95/07111

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	CROOKE ET AL., "Antisense Research and Applications," published 1993 by CRC Press (Boca Raton), pages 1-35, see pages 13-16.	1-51

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International application No.
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B. FIELDS SEARCHED
Electronic data bases consulted (Name of data base and where practicable terms used)

Analytical Abstracts, APS, BIOSIS, Derwent Biotechnology Abstracts, Chemical Abstracts, Cancer Literature Online, CJACS, Dissertation Abstracts, Derwent Drug File, EMBASE, GENBANK, Life Sciences Collection, Derwent World Patents Index

search terms: raf, A-raf, B-raf, C-raf, raf-1, protein kinase, oligonucleotide, antisense, cancer, proliferation, inhibition, transcription, translation, expression, Monia, Brett P., Boggs, Russell T.

sequence data bases searched: EST, GENBANK 89, GENBANK-NEW6, U-EMBL_89, EMBL-NEW6

sequence IDs searched: 2, 6, 12, 17, 20-27, 29, 37, and 40